

Identification of 7 α -Hydroxycephalosporin C as an Intermediate in the Methoxylation of Cephalosporin C by a Cell Free Extract of *Streptomyces clavuligerus*

John D. Hood,* Anna Elson, Martin L. Gilpin, and Allan G. Brown

Beecham Pharmaceuticals Research Division, Brockham Park, Betchworth, Surrey RH3 7AJ, U.K.

Incubation of cephalosporin C with a cell free extract of *Streptomyces clavuligerus* resulted in the formation of 7 α -hydroxycephalosporin C; the same crude enzyme preparation was shown to methylate synthetic 7 α -hydroxycephalosporin C to form 7 α -methoxycephalosporin C.

Streptomyces clavuligerus (ATCC 27064) produces the β -lactam antibiotic cephamycin C [7β -(5-amino-5-carboxyvaleramido)-3-carbamoyloxymethyl-7 α -methoxy-3-cephem-4-carboxylic acid]. The methoxy group has been shown to be derived from molecular oxygen and methionine.^{1,2} Furthermore, O'Sullivan and Abraham³ using a cell free extract of *S. clavuligerus* demonstrated the introduction of a methoxy group into certain cephalosporins.

In this communication we present evidence for the methoxylation of cephalosporin C (1) by an extract of *S. clavuligerus* proceeding by a two-step reaction involving the formation of 7 α -hydroxycephalosporin C (2) with subsequent methylation to yield 7 α -methoxycephalosporin C (3).

The preparation of a cell free extract and the reaction conditions for methoxylation of cephalosporin C were essentially as described by O'Sullivan and Abraham.³ The reaction was stopped with acetic acid and the supernatant (1 ml) was applied to a QAE Sephadex A25 column (0.5 \times 2 cm). The products were eluted with 0.2 M NaCl (1 ml) and submitted to h.p.l.c. Peaks (Figure 1) were detected at 9.0 min and 7.6 min corresponding to the retention times for synthetic 7 α -methoxycephalosporin C (3)⁴ and 7 α -hydroxycephalosporin C (2), respectively. Neither peak was present when samples were treated with cephalosporinase (prepared from *Enterobacter cloacae* McDermott) prior to chromatography.

To confirm the identification of 7 α -hydroxycephalosporin C (2) the oxygenation reaction was performed on a prepara-

tive scale from cephalosporin C (50 mg), using the described reaction conditions³ but excluding *S*-adenosylmethionine. Purification on QAE Sephadex and HP20 columns yielded a white solid (2.6 mg) which was identical [by fast atom bombardment (f.a.b.) mass, n.m.r., and u.v. spectroscopy, and h.p.l.c.] to the synthetic hydroxycephem (2).

Methylation of synthetic 7 α -hydroxycephalosporin C (2) was demonstrated using the same enzyme preparation and the reaction mixture contained 7 α -hydroxycephalosporin C (2)

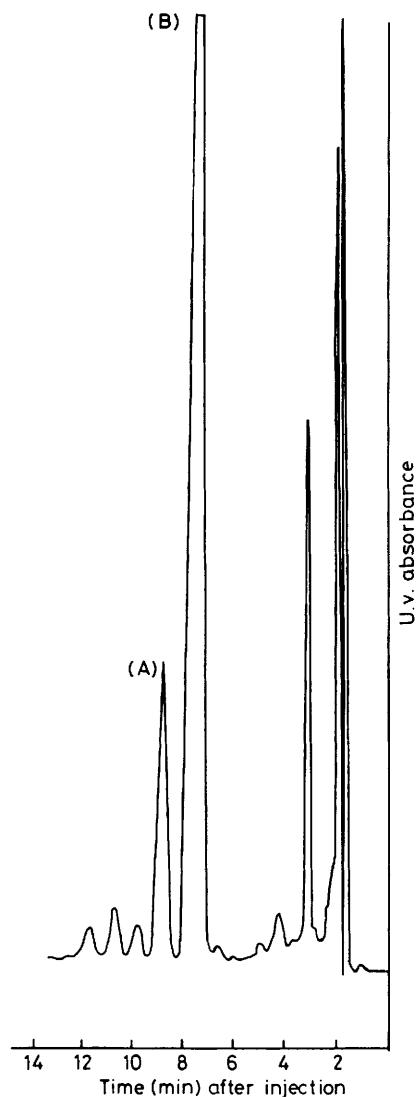
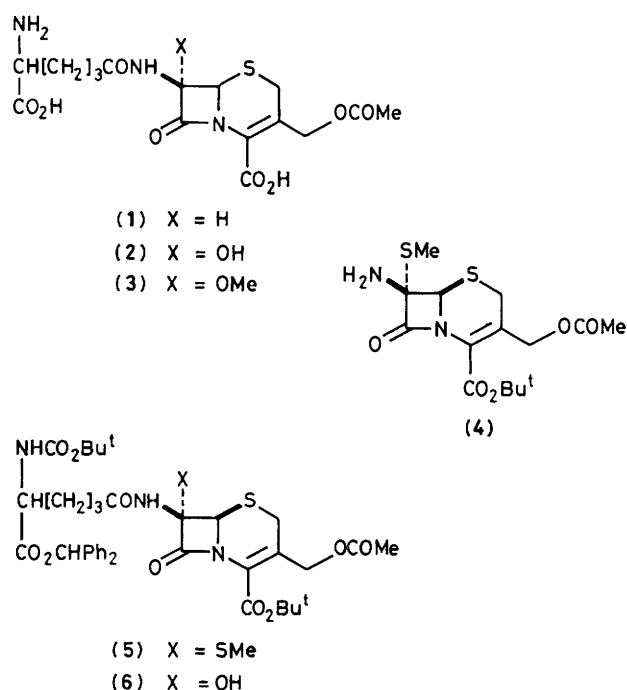


Figure 1. H.p.l.c. trace of the reaction products. H.p.l.c. conditions: C_{18} μ -Bondapak column eluted with 0.1 M NaH_2PO_4 , pH 4.2 at 2 ml min^{-1} . U.v. monitor at 260 nm. (A) 7-Methoxycephalosporin C; (B) 7-hydroxycephalosporin C.

(1 mM), *S*-adenosylmethionine (1 mM) in MOPS [= 3-(*N*-morpholino)propanesulphonic acid] buffer (0.05 M, pH 7.5). 7 α -Methoxycephalosporin C (3) was detected by h.p.l.c. (Figure 1).

7 α -Hydroxycephalosporin C (2) was prepared as follows: the amine (4)⁵ was acylated with the acid chloride derived from diprotected *D*- α -aminoadipic acid⁶ and the resulting 7 α -methylthiocephalosporin (5) was converted into the 7 α -hydroxy derivative (6) by treatment with mercury(II) acetate in aqueous tetrahydrofuran. Deprotection of the ester (6) with trifluoroacetic acid–anisole and subsequent neutralisation afforded the potassium salt of 7 α -hydroxycephalosporin C (2) {[α]_D²⁰ + 102° (*c* 1 in H₂O); ν_{\max} (KBr) 1760, 1660, and 1600 cm⁻¹; ¹H n.m.r. (D₂O) δ 1.7–1.9 (4H, overlapping m, CHCH₂CH₂), 2.08 (3H, s, CH₃CO₂), 2.42 (2H, t, *J* 7 Hz, CH₂CON), 3.29 and 3.65 (2H, ABq, *J* 17 Hz, 2-H), 3.73

(1H, t, *J* 7 Hz, CHNH), 4.65 and 4.81 (2H, ABq, *J* 13 Hz, CH₂OAc), and 5.10 (1H, s, 6-H); λ_{\max} (H₂O) 260 nm (ϵ 7 200); positive ion f.a.b. mass spectrum, *MH*⁺ 470, *MK*⁺ 508}.

Received, 13th June 1983; Com. 777

References

- 1 J. O'Sullivan, R. T. Aplin, C. M. Stevens, and E. P. Abraham, *Biochem. J.*, 1979, **179**, 47.
- 2 J. G. Whitney, D. R. Brannon, J. A. Mabe, and K. J. Wicker, *Antimicrob. Agents Chemother.* 1972, **1**, 247.
- 3 J. O'Sullivan and E. P. Abraham, *Biochem. J.*, 1980, **186**, 613.
- 4 G. A. Koppel and R. A. Kochler, *J. Am. Chem. Soc.*, 1973, **95**, 2403.
- 5 T. Jen, J. Frazee, and J. R. E. Hoover, *J. Org. Chem.*, 1973, **38**, 2857.
- 6 S. Wolfe and M. G. Jokinen, *Can. J. Chem.*, 1979, **57**, 1388.